Total Synthesis of (+)-Sch 725680: Inhibitor of Mammalian A-, B-, and Y-Family DNA Polymerases

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The total synthesis of (+)-Sch 725680, a member of the hydrogenated azaphilone family, has been accomplished. The synthesis confirmed the absolute configuration and biological activities of the natural product. A key reaction to construct a hydrogenated azaphilone core skeleton is a Ti-mediated aldol reaction.

Sch 725680, **1**, was originally isolated by Yang et al. from a culture of an *Aspergillus* sp. and was shown to display growth inhibitory activity against *Saccharomyces cerevisiae* PM503 and *Candida albicans* C43 with MICs of 8 and $64 \mu g/mL$, respectively (Figure 1).¹ Recently, Stierle et al. reisolated (+)-**1**, named berkazaphilone C, together with berkazaphilone B, **2**, from a culture of *Penicillium rubrum* as caspase 1 inhibitors with an IC₁₀₀ of 25 μ M.² They also reported that **1** selectively inhibits cell proliferation against the SR cell line in the National Cancer Institute (NCI) 60 cell line assay with a GI₅₀ of 0.38 μ M. In our natural product isolation program for the construction of a small molecule library,³ we also independently reisolated (+)-1 together with **2**, named pinophilin A, and pinophilin B, **3**, from a culture of *Penicillium pinophilum* Hedgcok as inhibitors of the mammalian A-, B-, and Y-family of DNA polymerases.^{3g} Using the excitation chirality method, we also determined that the absolute configurations are 7*S*,8*S*,8a*S*.

Compounds 1, 2, and 3 belong to a class of natural products called hydrogenated azaphilones, a subclass of azaphilones, that share a highly oxygenated bicyclic core, a

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⁽¹⁾ Yang, S.-W.; Chan, T.-M.; Terracciano, J.; Patel, R.; Patel, M.; Gullo, V.; Chu, M. J. Antibiot. 2006, 59, 720–723.

⁽²⁾ Stierle, A. A.; Stierle, D. B.; Girtsman, T. J. Nat. Prod. 2012, 75, 344–350.



Figure 1. Structure of Sch 725680, Berkazaphilone B, and Pinophilin B.

chiral tertiary hydroxyl group, and a trienone moiety.⁴ Although a number of synthetic studies of azaphilones, such as mitorubrinic acid,⁵ have been reported, no synthetic study of hydrogenated azaphilones has been disclosed. A flexible and scalable synthetic route for structure–activity relationship studies is required for further biological studies of hydrogenated azaphilones. Herein, we report the first total synthesis of (+)-1, as well as confirmation of its absolute configuration and inhibitory activity against the mammalian A-, B-, and Y-family of DNA polymerases and the proliferation of human cancer cell lines.

Our retrosynthesis of (+)-Sch 725680 is depicted in Scheme 1. Sch 725680, **1**, should be derived from hydrogenated azaphilone core structure **4** by the attachment of an orsellinate fragment. The hydrogenated azaphilone core skeleton should be prepared from diketone **5** using intramolecular aldol cyclization and 1,6-oxy-Michael cyclization. Diketone **5** should be derived from lactone **6**, which should be stereoselectively obtained from readily available imide 7^6 and aldehyde 8^7 using the Ti-mediated aldol reaction developed by Kobayashi's group.⁸





The Ti-mediated aldol reaction of imide 7 and aldehyde 8 (LDA, ClTi(O-*i*-Pr)₃, THF, 81%) gave anti-1,2-diol 9,⁹ followed by the deprotection of isopropylidene acetal and cleavage of the chiral auxiliary group $(Zn(NO_3)_2 \cdot 6H_2O_1)$ MeCN, 89%) to afford lactone 10 as a diastereomeric mixture of the hydroxy methyl group ($\alpha/\beta = 4:1$). Isopropylidene acetal protection of 1,3-diol 10 ((MeO)₂CMe₂, 10-camphorsulfonic acid (CSA), CH₂Cl₂) gave lactone 6α and 6β in yields of 70% and 18%, respectively. After both lactone 6α and 6β were converted to Weinreb amides (MeO(Me)NH·HCl, i-PrMgBr, THF), oxidation of alcohols (PCC, MS4A, CH₂Cl₂) gave corresponding aldehydes 11 β and 11 β in yields of 65% and 68% over two steps, respectively. The stereochemistry of the C8a formyl group in aldehyde 11β was inverted under basic conditions (DBU, THF, 96%) to afford aldehyde 11α , as shown in Scheme 2.

Freshly prepared dibromoalkene 12^{10} was treated with *n*-BuLi (THF, -78 °C to rt, 2 h in the dark) to give alkynyl lithium 13; the resulting solution was immediately used in the next step (Scheme 3). The treatment of aldehyde 11α with alkynyl lithium 13 followed by treatment with MeLi and oxidation of the resultant alcohol gave diketone 5. Aldol cyclization¹¹ of diketone 5 (TBAF, THF, 0 °C, 2 h) gave β -hydroxy ketone 14 and α , β -unsaturated ketone 15

⁽⁹⁾ The absolute configuration of secondary alcohol was determined as S by modified Mosher's method using (S)- and (R)-MTPA esters of 9. The relative configuration of 1,2-diol was determined as the *anti* configuration by NMR studies of acetonide derivatives 6α and 6β . See Supporting Information.



(10) (a) Bestmann, H. J.; Frey, H. *Liebigs Ann. Chem.* 1980, *12*, 2061–2071. (b) Bialy, L.; Waldmann, H. *Chem.*—*Eur. J.* 2004, *10*, 2759–2780.
(11) Mischne, M. *Tetrahedron Lett.* 2003, *44*, 5823–5826.

^{(3) (}a) Ogawa, A.; Murakami, C.; Kamisuki, S.; Kuriyama, I.; Yoshida, H.; Sugawara, F.; Mizushina, Y. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 3539–3543. (b) Kamisuki, S.; Ishimaru, C.; Onoda, K.; Kuriyama, I.; Ida, N.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *Bioorg. Med. Chem.* **2007**, *15*, 3109–3114. (c) Naganuma, M.; Nishida, M.; Kuramochi, K.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *Bioorg. Med. Chem.* **2008**, *16*, 2939–2944. (d) Nishida, M.; Ida, N.; Horio, M.; Takeuchi, T.; Kamisuki, S.; Murata, H.; Kuramochi, K.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *Bioorg. Med. Chem.* **2008**, *16*, 5115– 5122. (e) Kimura, T.; Nishida, M.; Kuramochi, K.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *Bioorg. Med. Chem.* **2008**, *16*, 4594–4599. (f) Kimura, T.; Takeuchi, T.; Kumamoto-Yonezawa, Y.; Ohashi, E.; Ohmori, H.; Masutani, C.; Hanaoka, F.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *Bioorg. Med. Chem.* **2008**, *16*, 4594–4599. (f) Kimura, D.; Takeuchi, T.; Kuramochi, K.; Kuriyama, I.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *Biosci. Biotechnol. Biochem.* **2010**, *74*, 793–801. (g) Myobatake, Y.; Takeuchi, T.; Kuramochi, K.; Kuriyama, K.; Ishido, T.; Hirano, K.; Sugawara, F.; Yoshida, H.; Mizushina, Y. *J. Nat. Prod.* **2012**, *75*, 135–141.

⁽⁴⁾ Osmanova, N.; Schultze, W.; Ayoub, N. Phytochem. Rev. 2010, 9, 315–342.

^{(5) (}a) Chong, R.; Gray, R. W.; King, R. R.; Whalley, W. B. J. Chem. Soc., Chem. Commun. **1970**, 101. (b) Whalley, W. B.; Chong, R.; Gray, R. W.; King, R. R. J. Chem. Soc. C **1971**, 3571–3575. (c) Marsini, M. A.; Gowin, K. M.; Pettus, T. R. R. Org. Lett. **2006**, *8*, 3481–3483. (d) Zhu, J.; Porco, A., Jr. Org. Lett. **2006**, *8*, 5169–5171.

^{(6) (}a) Murata, Y.; Kamino, T.; Aoki, T.; Hosokawa, S.; Kobayashi,
S. Angew. Chem., Int. Ed. 2004, 43, 3175–3177. (b) Onodera, Y.; Suzuki,
T.; Kobayashi, S. Org. Lett. 2011, 13, 50–53.

⁽⁷⁾ Bates, H. A.; Farina, J.; Tong, M. J. Org. Chem. 1986, 51, 2637-2641.

^{(8) (}a) Kamino, T.; Murata, Y.; Kawai, N.; Hosokawa, S.; Kobayashi, S. *Tetrahedron Lett.* **2001**, *42*, 5249–5252. (b) Murata, Y.; Kamino, K.; Hosokawa, S.; Kobayashi, S. *Tetrahedron Lett.* **2002**, *43*, 8121–8123.

Scheme 2. Preparation of Aldehyde 11α



at yields of 69% and 11% as ca. 10:1 and ca. 3:1 mixtures of E/Z isomers, respectively. Although isomerization of *E*-olefin to *Z*-olefin occurred, 12 we thought *Z*-olefin should be isomerized to *E*-olefin under 1.6-oxy-Michael cyclization conditions. Dehydration of alcohol 14 (Burgess reagent, toluene, 100 °C) gave enone 15 in 94% yield as a ca. 3:1 mixture of E/Z isomers. Isopropylidene acetal deprotection gave alcohol 16 (CSA, MeOH, 77% as a ca. 0.8:1 mixture of E/Z isomers) to cause further E/Z-isomerization. 1.6-Oxy-Michael cyclization of alcohol 16 was achieved using silver trifluoromethanesulfonate¹³ in CH₂Cl₂ to give hydrogenated azaphilone core skeleton 4 with only an E-olefin configuration, as expected. After benzyl deprotection (BCl₃, CH₂Cl₂, 0 °C, 67%) and the introduction of protected orsellinate (4-benzyloxy-2-methoxy-6-methylbenzoyl chloride, DMAP, i-Pr₂NEt, CH₂Cl₂, 53%),^{5d} both benzyl and methyl deprotection (BCl₃, CH₂Cl₂, 63%)^{5d} gave hydrogenated azaphilone Sch 725680, 1. The characterization data (¹H and ¹³C NMR, IR, MS, UV, CD, inhibitory activity of mammalian DNA polymerases and of the proliferation of human cancer and normal cells¹⁴) for 1 ($[\alpha]_D^{22}$ +103 (*c* 0.1, MeOH)) was in

(12) Ramage, R.; Sattar, A. Chem. Commun. 1970, 173-175.

agreement with those reported for $1^{1,2,3g}$ ([α]_D²² +103 (c 0.27, MeOH)^{3g}).





In summary, we have accomplished the first total synthesis of hydrogenated azaphilone Sch 725680, thereby, confirming its absolute configuration and biological activities. The natural product was obtained in a 13-step sequence with an overall yield of 6.0%, starting from known aldehyde **8**. Key steps to establish the hydrogenaged azaphilone core skeleton were a Ti-mediated aldol reaction with controlled introduction of the C7- and C8-stereochemistry and a silver-catalyzed 1,6-oxy-Michael cyclization.

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Supporting Information Available. Detailed experimental procedures, characterization data, ¹H and ¹³C NMR spectra, and biological activities of both synthetic and natural Sch 725680. This material is available free of charge via the Internet at http://pubs.acs.org.

^{(13) (}a) Wang, C.; Forsyth, C. J. Org. Lett. 2006, 8, 2997–3000.
(b) Nicolaou, K. C.; Cole, K. P.; Frederick, M. O.; Aversa, R. J.; Denton, R. M. Angew. Chem., Int. Ed. 2007, 46, 8875–8879.

⁽¹⁴⁾ Synthetic Sch 725680's ability to inhibit the activity of DNA polymerases and proliferation of five human cancer cell lines was investigated and compared to that of the natural substance. Synthetic Sch 725680 showed identical biological activity. See Supporting Information.

The authors declare no competing financial interest.